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① No organic Fluor cpnd used for Ser-phosphatase assays
② No Ser-linked Fluor. peptides
③ Fluor nonpeptides for alle phosphatase; tyrosine phosphatase assays.

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NEWS 17	Aug 08	PHARMAMarketLetter(PHARMAML) - new on STN
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NEWS 27	Oct 01	CASREACT Enriched with Reactions from 1907 to 1985
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=> s substrate (3a) (fluorescein or fluorescence or fluorescent)
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L4   ANSWER 1 OF 17  USPATFULL                              DUPLICATE 1
AN     2002:148557  USPATFULL
TI     KINASE ASSAYS USING POLYCATIONS
IN     NIKIFOROV, THEO T., 37 S. 16@H AVENUE, SAN JOSE  95112
PI     US 2002076697      A1   20020620
       US 6472141         B2   20021029
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AI US 2000-569193 A1 20000511 (9)
RLI Continuation-in-part of Ser. No. US 1999-316447, filed on 21 May 1999, GRANTED, Pat. No. US 6287774
PRAI US 1999-139562P 19990616 (60)
US 1999-156366P 19990928 (60)
DT Utility
FS APPLICATION
LREP CALIPER TECHNOLOGIES CORP, 605 FAIRCHILD DRIVE, MOUNTAIN VIEW, CA, 94043
CLMN Number of Claims: 62
ECL Exemplary Claim: 1
DRWN 25 Drawing Page(s)
LN.CNT 2153
CAS INDEXING IS AVAILABLE FOR THIS PATENT.
AB Methods, systems, kits for carrying out a wide variety of different assays that comprise providing a first reagent mixture which comprises a first reagent having a fluorescent label. A second reagent is introduced into the first reagent mixture to produce a second reagent mixture, where the second reagent reacts with the first reagent to produce a fluorescently labeled product having a substantially different charge than the first reagent. A polyion is introduced into at least one of the first and second reagent mixtures, and the fluorescent polarization in the second reagent mixture relative to the first reagent mixture is determined, this fluorescent polarization being indicative of the rate or extent of the reaction.

L4 ANSWER 2 OF 17 USPATFULL after F.D
AN 2002:227915 USPATFULL
TI Water-soluble, fluorescent, & electrophoretically mobile peptidic substrates for enzymatic reactions and methods for their use in high-throughput screening assays
IN Dwyer, Brian P., San Diego, CA, UNITED STATES
Havens, John R., San Diego, CA, UNITED STATES
PI US 2002123068 A1 20020905
AI US 2001-775840 A1 20010131 (9)
DT Utility
FS APPLICATION
LREP LYON & LYON LLP, 633 WEST FIFTH STREET, SUITE 4700, LOS ANGELES, CA, 90071
CLMN Number of Claims: 113
ECL Exemplary Claim: 1
DRWN 6 Drawing Page(s)
LN.CNT 2003
CAS INDEXING IS AVAILABLE FOR THIS PATENT.
AB The present invention provides non-radioactively labeled synthetic substrates for enzymatic reactions which exhibit markedly improved solubility having the general structure *F-R.sub.1-L.sub.1-R.sub.2-P.sub.Hc1-P.sub.S-P.sub.Hc2-(R.sub.3-L-R.sub.4-T).sub.y. These substrates may be designed to carry a charge to allow electrophoretic separation of substrates and reaction products. The invention also provides enzymatic activity assays for protein kinases, phosphatases and proteases utilizing the substrates of the invention, as well as methods of producing these substrates. In addition, the invention also provides libraries of the substrates, and methods of utilizing these libraries to select optimal synthetic peptide enzyme substrates for high-throughput screening assays.

L4 ANSWER 3 OF 17 USPATFULL
AN 2002:209306 USPATFULL
TI Methods of identifying nucleic acid sequences using polycations
IN Nikiforov, Theo T., San Jose, CA, United States
PA Caliper Technologies Corp., Mountain View, CA, United States (U.S. corporation)
PI US 6436646 B1 20020820
AI US 2000-727532 20001128 (9)

RLI Continuation of Ser. No. US 2000-569193, filed on 11 May 2000
Continuation-in-part of Ser. No. US 1999-316447, filed on 21 May 1999, now patented, Pat. No. US 6287774, issued on 11 Sep 2001
PRAI US 1999-139562P 19990616 (60)
US 1999-156366P 19990928 (60)
DT Utility
FS GRANTED
EXNAM Primary Examiner: Siew, Jeffrey
LREP Murphy, Matthew B.
CLMN Number of Claims: 12
ECL Exemplary Claim: 1
DRWN 31 Drawing Figure(s); 22 Drawing Page(s)
LN.CNT 1902
CAS INDEXING IS AVAILABLE FOR THIS PATENT.
AB Methods, systems, kits for carrying out a wide variety of different assays that comprise providing a first reagent mixture which comprises a first reagent having a fluorescent label. A second reagent is introduced into the first reagent mixture to produce a second reagent mixture, where the second reagent reacts with the first reagent to produce a fluorescently labeled product having a substantially different charge than the first reagent. A polyion is introduced into at least one of the first and second reagent mixtures, and the fluorescent polarization in the second reagent mixture relative to the first reagent mixture is determined, this fluorescent polarization being indicative of the rate or extent of the reaction.

L4 ANSWER 4 OF 17 CAPLUS COPYRIGHT 2002 ACS
AN 2001:618190 CAPLUS
DN 135:207447
TI Fluorescent assay for protein tyrosine phosphatases
IN Flint, Andrew J.; Cool, Deborah E.
PA Ceptyr, Inc., USA
SO PCT Int. Appl., 79 pp.
CODEN: PIXXD2
DT Patent
LA English
FAN.CNT 1

	PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
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	WO 2001061031	A3	20020307		
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	US 2002009762	A1	20020124	US 2001-788626	20010213
PRAI	US 2000-181769P	P	20000214		

AB The invention relates in part to screening assays for identifying agents that alter the interaction between a protein tyrosine phosphatase (PTP) and its tyrosine phosphorylated polypeptide substrate, using fluorescence energy signals generated by detectably labeled substrates. Assays are provided in certain embodiments, including high throughput screening assays, wherein candidate agents are screened by fluorescence polarization for their ability to influence (i) binding of substrate trapping mutant PTPs to substrates, or (ii) dephosphorylation of tyrosine phosphorylated substrates by PTPs.

L4 ANSWER 5 OF 17 USPATFULL
AN 2001:184724 USPATFULL

TI Methods and compositions for conducting processes in microfluidic devices
IN Wainright, Ann K., Cupertino, CA, United States
Visser, Irene, San Ramon, CA, United States
Singh, Sharat, San Jose, CA, United States
PA Aclara BioSciences, Inc., Mountain View, CA, United States (U.S. corporation)
PI US 6306273 B1 20011023
AI US 1999-291169 19990413 (9)
DT Utility
FS GRANTED
EXNAM Primary Examiner: Warden, Jill; Assistant Examiner: Starsiak, Jr., John S.
LREP Perkins Coie LLP
CLMN Number of Claims: 7
ECL Exemplary Claim: 1
DRWN 10 Drawing Figure(s); 10 Drawing Page(s)
LN.CNT 1849
AB A method is disclosed for controlling the direction and transport of a material on a microfluidic device formed from an acrylic polymer, by electrokinetic flow of a fluid containing the material. The method comprises providing an electrokinesis buffer containing a charged hydrophilic polymer, wherein charges on the polymer are randomly distributed. The method has applications for improved transport of proteins and transport of materials comprising differentially charged chemical species. A preferred embodiment of the present invention comprises use of the disclosed method for electrokinetic separations of a mixture of polypeptides having both positive and negative charges. Another preferred embodiment concerns use of the disclosed method for performing an assay involving mixing of two or more reagents, wherein a first reagent comprises an enzyme and a second reagent comprises species having an opposite charge to the enzyme.

L4 ANSWER 6 OF 17 ANABSTR COPYRIGHT 2002 RSC DUPLICATE 2
AN 64(19):F125 ANABSTR
TI An **assay** for phosphoinositide **phosphatases** utilizing **fluorescent substrates**.
AU Taylor, G. S.; Dixon, J. E.* (jedixon@umich.edu, Dept. Biol. Chem., Univ. Michigan Med. School, Ann Arbor, MI 48109-0606, USA).
SO Anal. Biochem. (2001) 295(1), 122-126
CODEN: ANBCA2 ISSN: 0003-2697
DT Journal
LA English
AB Incubation mixtures contained recombinant phosphatases myotubularin (MTM1) or the PTEN described previously (Maehama and Dixon, J. Biol. Chem., 1998, 273, 13375) at pH 6 and 8, respectively, and the fluorescent substrate di-C6-NBD6 phosphoinositide or its 3,4,5-triphosphate (Echelon). The reactions were stopped by adding acetone, the mixtures were evaporated and the residue was suspended in acetic acid/propan-2-ol/methanol (2:5:5). Portions were analysed by TLC on K6 silica gel with a mobile phase of water/acetic acid/acetone/methanol/chloroform (2:2:2:5:5), with evaluation by UV irradiation in an imaging workstation. The semi-quantitative method complemented the more quantitative malachite green method and was recommended for enzyme specificity studies.

L4 ANSWER 7 OF 17 BIOSIS COPYRIGHT 2002 BIOLOGICAL ABSTRACTS INC.DUPLICATE 3
AN 1998:313363 BIOSIS
DN PREV199800313363
TI Synthesis of **fluorescent substrates** for protein tyrosine **phosphatase assays**.
AU Watanabe, Takumi; Imoto, Masaya; Taniguchi, Hiroki; Kinoshita, Kazuhiko, Jr.; Umezawa, Kazuo (1)
CS (1) Inst. Microbial Chem., 3-14-23 Kamiosaki, Shinagawa-ku, Tokyo 141

Japan
SO Bioorganic & Medicinal Chemistry Letters, (June 2, 1998) Vol. 8, No. 11, pp. 1301-1302.
ISSN: 0960-894X.
DT Article
LA English
AB Two fluorescent substrates for protein tyrosine phosphatase (PTPase) reaction were prepared by conjugation of commercially available O-phosphotyrosine and dansyl chlorides. They were hydrolyzed by CD45 tyrosine phosphatase, and proved to be useful for PTPase assay.

L4 ANSWER 8 OF 17 BIOSIS COPYRIGHT 2002 BIOLOGICAL ABSTRACTS INC.DUPLICATE 4
AN 1995:347224 BIOSIS
DN PREV199598361524
TI Use of a novel human immunodeficiency virus type 1 reporter virus expressing human placental alkaline phosphatase to detect an alternative viral receptor.
AU He, Jianglin; Landau, Nathaniel R. (1)
CS (1) Aaron Diamond AIDS Res. Cent., 455 First Ave., New York, NY 10016 USA
SO Journal of Virology, (1995) Vol. 69, No. 7, pp. 4587-4592.
ISSN: 0022-538X.
DT Article
LA English
AB We report here on the construction and use of a novel human immunodeficiency virus (HIV) type 1 reporter vector, HIV-AP, that encodes human placental alkaline phosphatase. Upon staining with chromogenic alkaline phosphatase substrates 24 to 36 h postinfection, cells infected with HIV-AP develop an intense purple color and can then be counted under a dissecting microscope. Alternatively, HIV-AP infectivity can be quantitated and infected cells can be sorted by a fluorescence-activated cell sorter after staining with a **fluorescent** alkaline **phosphatase substrate**. The **assay** is rapid and accurate, has very low background in a variety of cell lines and primary cells, and is not restricted to use in human cells. Infectious HIV-AP can be pseudotyped by various HIV or murine leukemia virus envelope glycoproteins. Using this virus, we have addressed the long-standing question of CD4-independent infection of cells by HIV. Our results confirm the presence on a human osteosarcoma cell line of an alternative receptor for HIV infection that functions with an efficiency approximately 1/20 that of CD4.

L4 ANSWER 9 OF 17 BIOSIS COPYRIGHT 2002 BIOLOGICAL ABSTRACTS INC.DUPLICATE 5
AN 1993:498702 BIOSIS
DN PREV199396122709
TI Synthesis and **fluorescence** properties of a **substrate** for a continuous fluorimetric **assay** of protein tyrosine **phosphatases**.
AU Garcia-Echeverria, Carlos (1); Rich, Daniel H.
CS (1) Pharmaceutical Div., Ciba-Geigy Ltd., CH-4002 Basel Switzerland
SO Bioorganic & Medicinal Chemistry Letters, (1993) Vol. 3, No. 8, pp. 1601-1604.
ISSN: 0960-894X.
DT Article
LA English
AB A phosphotyrosine containing peptide, H-Thr-Glu-Pro-Glu-Tyr(PO-3H-2)-Gln-Pro-Gly-Glu-NH-2, has been synthesized to study the effect of phosphate substitution on the intrinsic fluorescence of the free tyrosine-peptide. The observed difference in fluorescence between the phosphorylated and unphosphorylated peptide at 300 nm upon excitation at 275 nm can be applied for continuous fluorimetric assays of protein-tyrosine phosphatases.

L4 ANSWER 10 OF 17 CAPLUS COPYRIGHT 2002 ACS
AN 1992:421221 CAPLUS
DN 117:21221
TI DNA hybridization assay using ATTOPHOS, a fluorescent substrate for alkaline phosphatase
AU Cano, Raul J.; Torres, Maria J.; Klem, Robert E.; Palomares, Jose C.
CS Biol. Sci. Dep., California Polytech. Univ., Obispo, CA, 93407, USA
SO BioTechniques (1992), 12(2), 264-6, 268-9
CODEN: BTNQDO; ISSN: 0736-6205
DT Journal
LA English
AB A fluorometric procedure for the detection of DNA-DNA hybrids is described. The procedure involved the detection of probe-bound alk. phosphatase with the fluorescent substrate ATTOPHOS. This substrate is converted to ATTOFLUOR by alk. phosphatase and fluoresces strongly at 550 nm when excited with a wavelength fo 440 nm. DNA hybridization assays were performed both with dilns. of purified target plasmid DNA (pSE9 or PBR322) and whole bacterial cells. Streptavidin-alk. phosphatase conjugates were added to react with bound probe. Fluorometric assays, as well as colorimetric assays, using 5-bromo-4-chloro-3-indolylphosphate + nitroblue tetrazolium for alk. phosphatase activity were performed. The fluorescence of the substrate was measured at time intervals, and the slope of the regression line calcd. A slope four times greater than that of background was considered pos. One hundred femtograms or 2.2 .times. 104 mols. of homologous DNA were detected with the fluorescent assay as compared with 10000 fg or 2.2 .times. 106 mols. of homologous DNA with the colorimetric assay. Similar results were obtained with whole cells. Approx. 1 .times. 103 homologous cells were detected fluorometrically and 1 .times. 105 cells were detected colorimetrically. Based on these results, the DNA hybridization assay described here using ATTOPHOS as the substrate for alk. phosphatase is a very sensitive assay for the detection of DNA-DNA hybrids.

L4 ANSWER 11 OF 17 CAPLUS COPYRIGHT 2002 ACS
AN 1991:651633 CAPLUS
DN 115:251633
TI Nucleic acid determination by phosphatase-labeled DNA probe using fluorescent coumarin compounds as the enzyme substrate
IN Fujita, Satoshi; Hori, Hiroshi; Shirahama, Haruhisa; Saito, Hiroshi
PA Aisin Seiki Co., Ltd., Japan
SO Jpn. Kokai Tokkyo Koho, 10 pp.
CODEN: JKXXAF
DT Patent
LA Japanese
FAN.CNT 1

PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
JP 03072898	A2	19910328	JP 1989-207203	19890810

OS MARPAT 115:251633
AB The title hybridization assay uses fluorescent coumarin compds. e.g. I [R1 = phosphate ester; X = hydroxy, Y = alkoxy] as the enzyme substrate. The method using I is sensitive and rapid and I are safe to use (i.e. compared to radiolabels). Thus, biotinylated DNA probe was hybridized with test DNA on a nitrocellulose membrane filter, treated with alk. phosphatase-avidin conjugate (com. kit), and then treated with a soln. contg. trimethoxybenzylcoumarin phosphate (prepn. given), NaCl, MgCl2, and Tris buffer at room temp. for 5 min-2 h. The reaction mixt. was fluorometrically measured for the DNA detn. The detection limit was 0.02 pg.

L4 ANSWER 12 OF 17 BIOSIS COPYRIGHT 2002 BIOLOGICAL ABSTRACTS INC.DUPLICATE 6
AN 1988:289985 BIOSIS
DN BA86:18252

TI QUANTITATIVE EVALUATION OF AXONAL REGENERATION BY IMMUNOCHEMICAL ASSAY FOR NEUROFILAMENT PROTEIN.
AU GUTH L; ALBERS R W; BARRETT C P; DONATI E J
CS DEP. ANATOMY, UNIV. MARYLAND SCH. MED., BALTIMORE, MARYLAND 21201.
SO EXP NEUROL, (1988) 100 (1), 83-97.
CODEN: EXNEAC. ISSN: 0014-4886.
FS BA; OLD
LA English
AB In experiments on nerve regeneration requiring assessment of the rate and extent of axonal outgrowth, the availability of a simple and accurate method of quantification would be extremely useful. We approached this issue by modifying the conventional ELISA procedure so as to provide a sensitive, specific, and quantitative biochemical assay of the phosphorylated neurofilament content of homogenates or sections of nerve tissue. The technique involves four sequential steps: (i) adhesion of fixed or fresh homogenates or tissue sections to wells of microtiter plates, (ii) binding of a monoclonal antibody against phosphorylated neurofilament to the tissue, (iii) secondary binding to the anti-phosphorylated neurofilament of a phosphatase-labeled second antibody (antimouse IgG), and (iv) enzymatic **assay** of alkaline **phosphatase** activity using a **fluorescent substrate** (4-methylumbelliferyl phosphate). The technique is sufficiently sensitive to measure the phosphorylated neurofilament content of a 1:100,000 (w/v) homogenate of brain, spinal cord, or peripheral nerve and of single 10-.mu.m paraffin sections of Bouin-fixed rat spinal cord. To validate the applicability of the procedure to the study of nerve regeneration, the sciatic nerve of adult rats was either crushed (to permit regeneration) or transected and ligated (to preclude regeneration). The animals were autopsied 1 to 16 weeks, later, when four segments 3-mm in length taken from regions proximal and distal to the lesion site were assayed for phosphorylated filament content. The temporal course of its disappearance during degeneration and its reappearance during regeneration coincided with the known histologic changes in crushed and transected nerves. These findings demonstrate the validity of using the immunochemical assay for PNF in studies of nerve regeneration in the peripheral nervous system and the potential applicability of this procedure to studies on regeneration in the central nervous system.

L4 ANSWER 13 OF 17 CAPLUS COPYRIGHT 2002 ACS
AN 1988:52016 CAPLUS
DN 108:52016
TI Hydrolyzable fluorescent substrates for phosphatases and analytical use thereof
IN Sundberg, Michael William; McClune, Gregory Joseph; Babb, Bruce Edward
PA Eastman Kodak Co., USA
SO Eur. Pat. Appl., 19 pp.
CODEN: EPXXDW
DT Patent
LA English
FAN.CNT 1

	PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
PI	EP 232129	A2	19870812	EP 1987-300775	19870129
	EP 232129	A3	19890830		
	R: CH, DE, FR, GB, LI				
	US 4803157	A	19890207	US 1986-824752	19860131
	CA 1273023	A1	19900821	CA 1986-507346	19860423
	JP 62190191	A2	19870820	JP 1987-18750	19870130
PRAI	US 1986-824752		19860131		

AB [BLOCK]XRfL [BLOCK] = phosphono or salts, thioxophosphono or salts; X = O, S or NR (R = H or (un)substituted alkyl; when [BLOCK] = thiophosphono, X = O; Rf= (un)substituted phenolenone or benzphenelenone moiety;L = H or a specific binding ligand; when released as XRfL, XRfL exhibits max. fluorescent emission at least 580 nm and max. absorption >530 nm] are

hydrolyzable **fluorescent substrates** for **phosphatases** and for **anal**. use. A sample contg. alk. phosphatase was treated with a reagent contg. phenalenone 6-phosphate, MgOAc, and pH 8.5 Tris buffer, and the reaction mixt. was measured at 580 nm with excitation at 535 nm for enzyme assay. Prepn. of phenalenone 6-phosphate is described.

L4 ANSWER 14 OF 17 CAPLUS COPYRIGHT 2002 ACS DUPLICATE 7
AN 1967:91824 CAPLUS
DN 66:91824
TI **Fluorescein** and naphthol **substrates** for **phosphatase assays**
AU Westley, John W.
CS Stanford Univ., Palo Alto, CA, USA
SO NASA (Nat. Aeronaut. Space Admin.) Access. (1964), NASA-CR-59380, 41 pp. Avail.: CFSTI, $3 hc
From: Sci. Tech. Aerospace Rept. 1965, 3(18), N65-29435
CODEN: NAACAF
DT Report
LA English
AB The chemistry of fluorescein, fluorescein derivs., and naphthol derivs. was investigated. Because of high quantum yields of fluorescence, fluorescein and related compds. provide a suitable starting point for the study of the problem of soil fluorescence which was observed in previous detns. of phosphatase activity in soils using fluorimetric assays. The high quantum yield of fluorescein means that 10-10 to 10-11M solns. can be detected using com. available fluorimeters. Bacillus subtilis was used to det. the bacterial hydrolysis of 6,6'-dihydroxynaphthofluoran diphosphate. Also, a comparison of the diphosphate and p-nitrophenyl phosphate as substrates for phosphatase is included.

L4 ANSWER 15 OF 17 CAPLUS COPYRIGHT 2002 ACS
AN 1965:411674 CAPLUS
DN 63:11674
OREF 63:2101d-e
TI Multivator, a biochemical laboratory for Martian experiments
AU Levinthal, E.; Hundley, L.; Lederberg, J.
CS Stanford Univ., Stanford, CA
SO Life Sci. Space Res. (1964), Volume Date 1963, (2), 112-23
DT Journal
LA English
AB The Multivator is a detection system for signs of life on Mars to be used in the constraints of Mariner-type missions. The basic instrument as well as several recent modifications are discussed and its operation after landing on Mars outlined in detail. The device will permit many different biochem. expts. to be carried out, particularly those which can use a photomultiplier as an output transducer and include fluorimetry, nephelometry, and scintillometry. The assays under study fall into two general categories: the first to detect the presence of hydrolytic enzymes by fluorimetry, and the second uses the techniques of membrane sepn. to measure production of mols. with membrane transmission varying from those of the **substrate**. Photometric **assays** of **fluorescence** of **phosphatase** have been given particular attention because of its distribution in terrestrial organisms and capability of detection.

L4 ANSWER 16 OF 17 CHEMINFORMRX COPYRIGHT 2002 FIZ CHEMIE
AN 199840216 CHEMINFORMRX
TI Synthesis of **Fluorescent Substrates** for Protein Tyrosine **Phosphatase Assays**.
AU WATANABE, T.; IMOTO, M.; TANIGUCHI, H.; KINOSHITA, K. JUN.; UMEZAWA, K.
CS Dep. Appl. Chem., Fac. Sci. Technol., Keio Univ., Kohoku, Yokohama 223, Japan
SO Bioorg. Med. Chem. Lett., 8(11), 1301-1302 (1998)

CODEN: BMCLE8 ISSN: 0960-894X
LA English
AB A new assay system for PTPase with fluorescent substrates (III) is developed.

L4 ANSWER 17 OF 17 BABS COPYRIGHT 2002 BEILSTEIN CDS MDLI
AN 6102223 BABS
TI Synthesis of **Fluorescent Substrates** for Protein Tyrosine **Phosphatase Assays**
AU Watanabe, Takumi; Imoto, Masaya; Taniguchi, Hiroki; Kinoshita, Kazuhiko; Umezawa, Kazuo
SO Bioorg.Med.Chem.Lett. (1998), 8(11), 1301-1302
CODEN: BMCLE8
DT Journal
LA English
SL English
AB Two fluorescent substrates for protein tyrosine phosphatase (PTPase) reaction were prepared by conjugation of commercially available O-phosphotyrosine and dansyl chlorides.They were hydrolyzed by CD45 tyrosine phosphatase, and proved to be useful for PTPase assay.

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=> s l2 (3a) peptide
  20 FILES SEARCHED...
  31 FILES SEARCHED...
  47 FILES SEARCHED...
  64 FILES SEARCHED...
  83 FILES SEARCHED...
L5          819 L2 (3A) PEPTIDE

=> s l5 (5a) l1
  28 FILES SEARCHED...
  52 FILES SEARCHED...
  72 FILES SEARCHED...
L6            1 L5 (5A) L1

=> d l6 bib ab

L6   ANSWER 1 OF 1  USPATFULL
AN     2002:227915  USPATFULL
TI     Water-soluble, fluorescent, & electrophoretically mobile peptidic
       substrates for enzymatic reactions and methods for their use in
       high-throughput screening assays
IN     Dwyer, Brian P., San Diego, CA, UNITED STATES
       Havens, John R., San Diego, CA, UNITED STATES
PI     US 2002123068      A1   20020905
AI     US 2001-775840     A1   20010131 (9)
DT     Utility
FS     APPLICATION
LREP   LYON & LYON LLP, 633 WEST FIFTH STREET, SUITE 4700, LOS ANGELES, CA,
       90071
CLMN   Number of Claims: 113
ECL    Exemplary Claim: 1
DRWN   6 Drawing Page(s)
LN.CNT 2003
CAS INDEXING IS AVAILABLE FOR THIS PATENT.
AB     The present invention provides non-radioactively labeled synthetic
       substrates for enzymatic reactions which exhibit markedly improved
       solubility having the general structure *F-R.sub.1-L.sub.1-R.sub.2-
       P.sub.Hc1-P.sub.S-P.sub.Hc2-(R.sub.3-L-R.sub.4-T).sub.y. These
       substrates may be designed to carry a charge to allow electrophoretic
       separation of substrates and reaction products. The invention also
       provides enzymatic activity assays for protein kinases, phosphatases and
       proteases utilizing the substrates of the invention, as well as methods
       of producing these substrates. In addition, the invention also provides
       libraries of the substrates, and methods of utilizing these libraries to
       select optimal synthetic peptide enzyme substrates for high-throughput
       screening assays.


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